

# A Model for the UV-irradiation of Eumelanin supports Dual Functionality at Solar UV-Intensity

Rachel M. Haywood and Claire Linge

RAFT Institute of Plastic Surgery, Mount Vernon Hospital, Northwood, Middlesex, HA6 2RN, UK.

Although it is well established that eumelanin can be both UV-photosensitiser and radical scavenger its biological role remains unclear. ESR detection of superoxide in UV-irradiated DL-dopa melanin was reported in 1978 [1]; however, Persad *et al.* could only detect superoxide radicals in melanins isolated from red but not black hair [2]. Studies to assess photodamage by UV-irradiated melanins conflict [3,4]; and measurement of hydrogen peroxide production suggests dopa melanin is a photosensitiser over a wide concentration range [5], contrasting an earlier study demonstrating scavenging properties of the pigment [6].

Radical detection is complicated by insolubility of the pigment after isolation by conventional methods; and instability of the radical-adduct of DMPO at pH 7. DMPO-O<sub>2</sub>H<sup>•</sup>, however, is readily detected at pH 4.5: since some evidence suggests that the melanosome is acidic; radical production at pH 4.5 and pH 7 is similar (UV absorption to produce semiquinone radicals that disproportionate or donate an electron to oxygen), radical production by UV-irradiated, soluble, DL- and L-dopa melanins at different concentrations was investigated using DMPO at pH 4.5. At a fluence comparable to solar UV, production of the hydroperoxyl radical-adduct increases initially with concentration until about 0.3 mg/ml before subsequently decreasing with further concentration increase. The concentrations of DMPO-O<sub>2</sub>H<sup>•</sup> (observed to be steady state) are consistent with those predicted by a simple model based upon UV absorption and radical scavenging; and which assumes semiquinone-radical reduction of oxygen is fast compared to disproportionation. The data supports a previously hypothesised biological dual role for melanins at solar UV fluence.

- [1] Felix, C.C., Hyde, J.S., Sarna, T. & Sealy R.C. *Biochem. Biophys. Res. Commun.* **84**, 335-341 (1978).
- [2] Persad, S., Aravindakshan, M. & Haberman, H.F. *Photochem. Photobiol.* **37**, 63-68 (1983).
- [3] Aravindakshan Menon, I., Persad, S., Ranadive, N.S. & Haberman H.F. *Cancer Res.* **43**, 3165-3169 (1983).
- [4] Schmitz, S. *et al.* *Photochem. Photobiol.* **61**, 650-655 (1995).
- [5] Korytowski, W., Pilas, B., Sarna, T. & Kalyanaraman, B. *Photochem. Photobiol.* **45**, 185-190 (1987).
- [6] Korytowski, W., Kalyanaraman, B., Menon, I.A., Sarna, T. & Sealy R.C. *Biochim. et Biophys. Acta.* **882**, 145-153 (1986).